

CONVERGENT EVOLUTION OF RED CAROTENOID COLORATION IN WIDOWBIRDS AND BISHOPS (*EUPLECTES* SPP.)

Maria Prager^{1,2} and Staffan Andersson¹

¹Department of Zoology, University of Gothenburg, Box 463, 405 30 Göteborg, Sweden

²E-mail: maria.prager@zool.gu.se

Received November 25, 2009

Accepted June 22, 2010

Avian carotenoid-based signals are classic examples of sexually selected, condition-dependent threat displays or mate choice cues. In many species, male dominance or mating success is associated with redder (i.e., longer wavelength) color hues, suggesting that red colors are either more efficient or more reliable signals than yellow colors. Few studies, however, have investigated selection for redness in a macroevolutionary context. Here, we phylogenetically reconstruct the evolution of carotenoid coloration in the African widowbirds and bishops (*Euplectes* spp.), for which agonistic selection for redder hues, as well as pigmentary mechanisms, is well documented. Using reflectance spectrometry for objective color quantification, and accounting for phylogenetic uncertainty, we find that yellow plumage color is a retained ancestral state in *Euplectes*, and that red color hues have convergently evolved two or three times. Results are discussed in relation to a known diversity in pigment mechanisms, supporting independent origins of red color, and suggesting that agonistic selection and physiological constraints have interacted to generate color diversity in *Euplectes*.

KEY WORDS: Honest signaling, maximum likelihood, Ploceidae, Ploceinae, spectral reflectance, stochastic character mapping.

Extravagant color displays in birds are familiar examples of sexually selected signals, functioning in competition over mates, via intrasexual contests or intersexual attractiveness (Andersson 1994; Hill and McGraw 2006a). In a phylogenetic perspective, however, many questions remain regarding the evolution and diversification of color signals.

Brilliant red, orange, and yellow colors are typically achieved by deposition of carotenoid pigments into feathers, skin, or beak tissue (Fox and Vevers 1960; Brush 1978; Stradi 1998). Like all animals, birds must acquire carotenoids exclusively through the diet (Brockmann and Völker 1934; Goodwin 1984; Hill and McGraw 2006b), and carotenoid-based displays are therefore potentially limited by food intake (Endler 1980; Hill 1990). Furthermore, the uptake and expression of carotenoids can be affected by parasites (Hamilton and Zuk 1982; Milinski and Bakker 1990; Thompson et al. 1997; Zahn and Rothstein 1999) as well as direct allocation conflicts with immunological or antioxidant systems

(Lozano 1994; von Schantz et al. 1999; Møller et al. 2000; Blount et al. 2003). For any or several of these reasons, striking carotenoid colors are presumed to indicate health or condition across a wide range of taxa (Olson and Owens 1998; McGraw 2006, but see, e.g., Hartley and Kennedy 2004).

In many carotenoid-pigmented birds (Shawcross and Slater 1983; Hansen and Rohwer 1986; Hill 1990; Evans and Hatchwell 1992; Wolfenbarger 1999; Pryke and Griffith 2006; Pryke 2007) and other vertebrates (Bakker and Sevenster 1983; Evans and Norris 1996), mating success or dominance is associated with redder (i.e., longer wavelength) color hues. As observed by Hill (1996), this suggests that red carotenoid-based signals generally are more efficient or more reliable (honest) than yellow signals (Schluter and Price 1993; Andersson 2000). For example, red signals may more efficiently exploit sensory or cognitive receiver biases (Endler and Basolo 1998), or be more honest quality advertisements due to higher production or maintenance costs (Hudon

1991; Hill 1996). In either case, exaggeration of carotenoid color signals toward redder hues is expected. This is true also for purely condition-dependent signals, given that senders evolve physiological or behavioral adaptations that reduce signaling costs (Hill 1994; Badyaev 2004).

In a macroevolutionary context, the above predicts directional and convergent evolution of red carotenoid coloration, but how can this be reconciled with the view of sexual selection as labile and divergent in many animals (Wiens 2001), including several bird groups (e.g., Kusmierski et al. 1997; Omland and Lanyon 2000; Kimball et al. 2001; Cardoso and Mota 2008; but see Prager and Andersson 2009)? Among these, only the study of New World orioles *Icterus* spp (Hofmann et al. 2006) concerned carotenoid coloration, and the lability of color observed in this genus was recently contrasted with convergent yellow to red evolution in the sister genus of caciques (*Cacicus*, Kiere et al. 2009). Further phylogenetic analyses of color evolution are clearly needed, especially in taxa where both signal selection and signal mechanisms are better understood.

Phylogenetic and comparative analyses of signal diversity may enable interesting inferences of the past evolution as well as the current function of carotenoid pigmentation. Ancestral character state reconstruction may reveal interesting macroevolutionary patterns, such as convergence and directional change in plumage signals (e.g., Omland and Lanyon 2000; Hofmann et al. 2006), and suggest key taxa for selection analyses and experiments (Omland and Hofmann 2006). This interaction between macro- and microevolutionary studies becomes a particularly powerful approach if also combined with data on the proximate mechanisms of coloration, which may help identify genetic and developmental constraints on signal evolution (Harvey and Pagel 1991). Knowledge of the proximate basis of coloration is also important for defining and scoring characters, as well as for selecting appropriate algorithms for ancestral state reconstruction (Hofmann et al. 2006). Yet, to date, few reconstructions of color evolution have been performed on taxa where the underlying pigment mechanisms have been analyzed in detail.

A genus for which both the adaptive function and proximate mechanisms of carotenoid coloration are unusually well documented are the African widowbirds and bishops (*Euplectes* spp.). These are sexually and seasonally dichromatic weaverbirds (subfam. Ploceinae), breeding semi-colonially and polygynously in grasslands of sub-Saharan Africa. Male nuptial plumages show remarkable interspecific variation in carotenoid color expression, ranging from the small yellow epaulettes of certain widowbirds to the extensive scarlet red body plumage of many bishop species. The bright carotenoid plumage patches are displayed against contrasting black melanin pigmentation, and are emphasized through posturing in agonistic and courtship interactions (Crook 1964). Behavioral observations and experiments in two phylogenetically

distant *Euplectes* species have furthermore shown that males with redder (longer wavelength) carotenoid hues are more successful in male–male competition over territories (Pryke et al. 2001, 2002; Pryke and Andersson 2003a,b). In contrast, the more or less extremely elongated nuptial tails of widowbird males are selected by female mate choice (Andersson 1982, 1992; Pryke and Andersson 2002, 2005).

The presence of carotenoids in bishop feathers was established already by Kritzler (1943), and plumage as well as plasma pigments have recently been analyzed in several *Euplectes* species (Andersson et al. 2007; Prager et al. 2009). The results indicate that yellow feather colors primarily derive from direct deposition of the dietary yellow carotenoids lutein and zeaxanthin. Red color signals are, on the contrary, achieved by two distinctly different biochemical strategies: either addition of relatively small amounts of metabolically derived red C4-keto-carotenoids (mainly α -doradoxanthin and canthaxanthin), or deposition of high concentrations of both dietary and derived yellow pigments (Andersson et al. 2007; Prager et al. 2009).

In this study, we use a subspecies-level phylogeny of *Euplectes* (Prager et al. 2008) and spectral reflectance data to reconstruct and explore two aspects of the evolution of avian carotenoid coloration: First, the much more common occurrence of yellow than red plumage colors, across the weaverbirds (Ploceinae, ca. 120 species in 17 genera), may indicate that red is a more derived state. If supported by rigorous ancestral state reconstruction, it would suggest directional evolution of the presumably more costly and honest red carotenoid displays (Hill 1996). Second, the two distinct “red mechanisms” that we have identified in *Euplectes* (Andersson et al. 2007) suggest convergent signal evolution. Like the convergent evolution of elongated tails in this genus (Prager and Andersson 2009), this would provide a contrast to the common view of sexual selection as labile and divergent (e.g., Wiens 2001). We test these hypotheses by objectively quantifying carotenoid color hues, and by phylogenetically reconstructing color evolution within the genus *Euplectes*, while accounting for uncertainty in phylogeny, and comparing results from different character coding schemes and reconstruction methods.

Methods

SAMPLES AND MEASUREMENTS

Carotenoid color hue was examined in 33 *Euplectes* taxa, representing all 17 *Euplectes* species, and eight outgroup taxa, all of which are included in the molecular phylogeny by Prager et al. (2008). Measurements were acquired from live birds in the field, and from skin specimens at the Natural History Museum at Tring, UK (Table 1). Although *Euplectes* color hues are generally homogeneous within and between brightly carotenoid-pigmented

Table 1 Plumage coloration in *Euplectes*, i.e., widowbirds (w.) and bishops (b.), and outgroup genera *Amblyospiza* (A.), *Foudia* (F), *Ploceus* (P), and *Quelea* (Q). Continuous hue (spectral position) was measured as the wavelength at which reflectance is halfway between its minimum and its maximum value (λ_{R50}). Discrete hue classes were derived by k-means clustering of taxon mean values, and complemented by a category for taxa lacking carotenoid pigmentation (non-car.), resulting in three- and four-state classifications, respectively.

Taxon Scientific name ¹	Common name	Origin	Patch	n	Continuous hue ($\lambda_{R50} \pm SE$)	Discrete hue category	
						3-state	4-state
<i>A. albifrons albifrons</i>	thick-billed weaver	–	–	–	–	non-car.	non-car.
<i>E. afer afer</i>	yellow-crowned b.	Cameroon & Nigeria ²	rump	10	520.3±1.3	yellow	yellow
<i>E. afer ladaensis</i>	yellow-crowned b.	S Sudan to N Tanzania ²	rump	9	522.1±1.8	yellow	yellow
<i>E. afer taha</i>	yellow-crowned b.	Wakkerstroem, South Africa	rump	13	527.4±0.6	yellow	yellow
<i>E. albonotatus albonotatus</i>	white-winged w.	E South Africa to Malawi ²	Iwc ³	10	525.7±1.3	yellow	yellow
<i>E. albonotatus asymmetricus</i>	white-winged w.	Sao Tomé, S.T. and Principe	Iwc	15	519.3±1.0	yellow	yellow
<i>E. albonotatus eques</i>	white-winged w.	–	–	–	–	non-car.	non-car.
<i>E. ardens ardens</i>	red-collared w.	Pietermaritzburg, South Africa	breast	34	568.0±1.7	red	orange
<i>E. ardens concolor</i>	red-collared w.	–	–	–	–	non-car.	non-car.
<i>E. ardens suahelicus</i>	red-collared w.	S Kenya ²	breast	10	595.2±2.7	red	red
<i>E. aureus</i>	golden-backed b.	Sao Tomé, S.T. and Principe	rump	10	535.7±2.1	yellow	yellow
<i>E. axillaris axillaris</i>	fan-tailed w.	Pietermaritzburg, South Africa	Iwc	14	590.0±2.4	red	red
<i>E. axillaris phoeniceus</i>	fan-tailed w.	SW Uganda ²	Iwc	10	586.3±1.4	red	red
<i>E. capensis approximans</i>	yellow-rumped w.	KwaZulu-Natal, South Africa ²	Iwc	10	519.8±0.8	yellow	yellow
<i>E. capensis crassirostris</i>	yellow-rumped w.	S Kenya ²	Iwc	10	521.2±0.7	yellow	yellow
<i>E. capensis phoenicomereus</i>	yellow-rumped w.	W Cameroon ²	Iwc	10	516.5±0.5	yellow	yellow
<i>E. diadematus</i>	fire-fronted b.	SE Kenya	forehead	3	584.0±9.0	red	red
<i>E. franciscanus E</i>	northern red b.	N Uganda ²	nape	4	603.8±5.4	red	red
<i>E. franciscanus W</i>	northern red b.	Nigeria to N Cameroon ²	nape	10	602.1±2.4	red	red
<i>E. gierowii ansorgei</i>	black b.	NE DRC to SW Ethiopia ²	breast	15	560.2±1.9	red	orange
<i>E. gierowii friederichseni</i>	black b.	N Tanzania ²	breast	4	575.0±3.8	red	orange
<i>E. harilaubi humeralis</i>	marsh w.	Entebbe, Uganda ²	Iwc	2	555.0±1.0	yellow	orange
<i>E. hordeaceus craspedopterus</i>	black-winged b.	Ethiopia ²	crown	10	589.8±2.1	red	red
<i>E. hordeaceus hordeaceus</i>	black-winged b.	Sao Tomé, S.T. and Principe	crown	10	585.9±3.6	red	red
<i>E. jacksoni</i>	Jackson's w.	–	–	–	–	non-car.	non-car.
<i>E. macrourus macrocerus N</i>	yellow-mantled w.	Ethiopia ²	Iwc	10	521.7±1.4	yellow	yellow
<i>E. macrourus macrocerus S</i>	yellow-mantled w.	Uganda to W Kenya ²	Iwc	5	513.6±2.2	yellow	yellow
<i>E. macrourus macrourus</i>	yellow-mantled w.	Nigeria ²	Iwc	10	519.7±1.0	yellow	yellow
<i>E. nigroventris</i>	zanzibar b.	E Tanzania ²	nape	10	584.9±1.8	red	red
<i>E. orix nigrifrons</i>	southern red b.	SW Uganda ²	crown	10	590.6±1.6	red	red
<i>E. orix orix</i>	southern red b.	Pietermaritzburg, South Africa	crown	22	580.1±1.7	red	red
<i>E. progné delamerei</i>	long-tailed w.	S Kenya ²	Iwc	5	591.8±3.7	red	red

Continued.

Table 1 Continued.

Taxon Scientific name ¹	Common name	Origin	Patch	<i>n</i>	Continuous hue ($\lambda_{R50} \pm SE$)	Discrete hue category	
						3-state	4-state
<i>E. progné</i>	long-tailed w.	NE South Africa ²	lwc	9	595.0±2.6	red	red
<i>E. psammocromius</i>	mountain marsh w.	S Highlands, Tanzania ²	lwc	5	519.3±1.9	yellow	yellow
<i>F. madagascariensis</i>	Madagascar red fody	Mahé, Seychelles	breast	18	582.7±2.2	red	red
<i>F. sechellarum</i>	Seychelles fody	Cousin Island, Seychelles	breast	3	528.2±3.1	yellow	yellow
<i>P. luteolus luteolus</i>	little weaver	Nigeria ²	breast	4	513.0±0.9	yellow	yellow
<i>P. melanocephalus fischeri</i>	black-headed weaver	Uganda ²	belly	6	513.8±3.5	yellow	yellow
<i>P. taeniopterus taeniopterus</i>	north. masked weaver	Sudan ²	breast	6	535.8±4.2	yellow	yellow
<i>Q. cardinalis</i>	cardinal quelea	Uganda to W Kenya ²	breast	5	598.8±2.2	red	red
<i>Q. quelea lathamii</i>	red-billed quelea	Transvaal, South Africa ²	breast	5	609.8±2.8	red	red

¹Taxonomic references are detailed in Prager et al. (2008).²Specimens from the Natural History Museum at Tring, UK.³Lesser wing coverts.

patches of individual male breeding plumages (Prager and Andersson, pers. obs.), two taxa (*E. diadematus* and *E. gierowii ansorgei*) have two differently colored carotenoid regions. Here, we measured and reconstructed the longest wavelength (i.e., “reddest”) feather hue displayed by breeding males of each taxon.

Depending on the proximate mechanisms underlying color expression, color hue can vary continuously, as well as group into more or less discrete classes, such as yellow, orange, or red. Assumptions of the mechanistic basis of interspecific color variation, combined with observations of current color distributions, have accordingly been used to justify either continuous (Hofmann et al. 2006) or discrete (Wang and Shaffer 2008) treatment of color hue in ancestral state reconstructions. Since high-performance liquid chromatography (HPLC) carotenoid profiles of *Euplectes* feathers (Andersson et al. 2007; Prager et al. 2009) suggest that evolutionary changes in color hue may have resulted from both quantitative and qualitative changes in pigmentation (see Introduction), we treat and reconstruct color hue both as a continuous and discrete trait in this study.

Continuous hue

Objective and context-independent color measures were acquired using reflectance spectrometry. Reflectance was measured with a USB2000 spectroradiometer system (OceanOptics, Dunedin, FL), including a fiber-optic reflectance probe, a HL2000 tungsten-halogen light source, and using CSPEC software (Ancal, Las Vegas, NV). We used a “coincident normal” measuring configuration (coaxial illumination and reading beams, perpendicular to the plumage plane), with a homemade probe holder fitted on the probe, taking three to five scans and removing the probe between each. The λ_{R50} colorimetric (wavelength at which reflectance is halfway between its minimum and its maximum) was used as a measure of the spectral location, that is, hue of plumage colors. For these and other details, see Andersson and Prager (2006).

Discrete hue

Following a procedure modified from Wang and Shaffer (2008), we used one-dimensional k-means clustering in the software “R” (R Development Core Team 2009) to partition the continuous λ_{R50} values into hue categories. “k-means clustering” (Hartigan and Wong 1979) is an iterative process in which *n* observations are partitioned into *k* clusters, while minimizing distances between sample points and cluster centroids. Initial values for centroids are randomly assigned, and new values are calculated as cluster means after each iteration, until cluster assignments no longer change. To find the most robust assignment of carotenoid hue classes in our dataset, three trials of k-means clustering with *k* = 2, *k* = 3, and *k* = 4, respectively, were each run on 1000 bootstrapped samples of taxon mean values. The percentage of runs in which

original data points would fall into the same (best) clusters was then compared between the three different trials.

In trials with $k = 2$ and $k = 3$, 100% of the runs returned the same clusters, loosely corresponding to the human-subjective classes “yellow” ($\lambda_{R50} = 513\text{--}555$ nm)/“red” (560–610 nm) and “yellow” (513–536 nm)/“orange” (555–575 nm)/“red” (580–610 nm), respectively. In contrast, clustering with $k = 4$ was less robust, giving identical results in only 43.8% of the runs. Ancestral reconstructions of discrete color hue were thus based on the former two classifications (i.e., $k = 2$ and $k = 3$). In order to include taxa lacking carotenoid pigmentation (and thus λ_{R50} value) in the analyses, a “non-carotenoid” class was added to each of these classification schemes (hereafter called “three-state” and “four-state” color hue).

RECONSTRUCTION OF COLOR EVOLUTION

We reconstructed continuous and discrete measures of color hue in *Euplectes* and outgroups.

Continuous hue

Reconstructions of continuous hue states were performed in the software Mesquite (Maddison and Maddison 2004) version 2.71, using the fully resolved (“allcompat”) version of the Bayesian consensus tree in Prager et al. (2009). Before reconstruction of continuous hue, alternative models of character evolution were compared using the Akaike Information Criterion (AIC; Akaike 1974) in the Mesquite module CoMET (Oakley et al. 2005; Lee et al. 2006), varying the punctuation asymmetry threshold between 5 and 1000. We found that color hue evolution in the genus is best described by a “punctuated avg/equal” model, although this is not significantly better ($\Delta\text{AIC} < 2$) than a “punctuated avg/distance” model. Both models limit trait change to one of each pair of descendants of a node, but whereas the former assumes that the amount of change between subsequent nodes is equal across the tree, the latter expects it to be proportional to genetic distance, or time, separating these nodes.

Among available options for reconstruction of continuously varying characters in Mesquite, that is, linear parsimony (Farris 1970; Swofford and Maddison 1987) and squared-change parsimony (Rogers 1984; Huey and Bennett 1987), we follow Hofmann et al. (2006) and Prager et al. (2009) in considering linear parsimony to be the best correspondence to the punctuated models suggested by CoMET. However, in order to test the sensitivity of our results to choice of method, we also reconstructed color hue evolution using squared-change parsimony.

Discrete hue

Discrete color hues of ancestral nodes were mapped onto the Bayesian consensus tree in Mesquite, using ordered maximum

parsimony and maximum-likelihood (Markov k-state 1 parameter model, Lewis 2001) methods. To account for uncertainty in the estimated tree, when making inferences about ancestral states, we also applied stochastic character mapping (Nielsen 2002; Huelsenbeck et al. 2003) of four-state color hue to the “post-burnin” posterior sample of 10,000 trees in Prager et al. (2009), using SIMMAP (Bollback 2006) version 1.0 beta 2.0.5.

Stochastic character mapping is a Bayesian reconstruction method, in which ancestral character states are sampled in proportion to their posterior probabilities, which are probabilities conditional on the observed distribution of states among extant taxa, and a stochastic model of character transformation. Markov chain Monte Carlo (MCMC) sampling of both alternative trees and model parameters, moreover, enables SIMMAP to calculate marginal posterior probabilities of each character state at selected ancestral nodes, i.e., probabilities that are integrated over alternative tree topologies, branch lengths, and transformation rates. For multistate characters, SIMMAP implements only a single-rate model for character state change, meaning that all state transitions are assumed to be equally likely, for a given branch length or time interval. To sample overall rate values (or “tree lengths”) in proportion to posterior probability, SIMMAP requires prior knowledge of the probability of different rate values to be specified, in the form of a discrete gamma (Γ) distribution. The gamma distribution is defined by its “shape” and “rate” parameters, α and β [mean, $E(\Gamma) = \alpha/\beta$, and standard deviation, $SD(\Gamma) = \sqrt{(\alpha/\beta^2)}$], and is discretized by SIMMAP using k categories.

Since rates of morphological evolution are generally estimated from data on a single or a few characters, rather than thousands of nucleotides in a typical DNA sequence alignment, these estimates are potentially more sensitive to choice of priors. Therefore, after scaling all tree lengths to one, we compared the results from four different priors on the overall transformation rate: (1) an empirical prior set to mimic the distribution of tree lengths derived from maximum likelihood reconstruction of discrete color hue on the posterior sample of trees in Mesquite ($\alpha = 100$, $\beta = 6.7$, i.e., $E[\Gamma] = 15$, $SD[\Gamma] = 1.5$); (2) a less informative prior with the same mean ($\alpha = 3$, $\beta = 0.2$, i.e., $E[\Gamma] = 15$, $SD[\Gamma] = 8.7$); (3) a prior of intermediate spread, with mean set to the observed low extreme of the likelihood distribution ($\alpha = 5$, $\beta = 0.5$, i.e., $E[\Gamma] = 10$, $SD[\Gamma] = 4.5$); and (4) a prior of intermediate spread, with mean set to the high extreme of the likelihood distribution ($\alpha = 10$, $\beta = 0.5$, i.e., $E[\Gamma] = 20$, $SD[\Gamma] = 6.3$). The gamma distribution was discretized using 50 categories, and the number of draws made from the prior distribution of rates was set to 50 per tree. Following each analysis, the posterior distribution of the rate parameter was inspected to make sure that it was not truncated at either end, thus warranting limits on the prior to be changed.

Results

SAMPLES AND MEASUREMENTS

Mean spectral reflectance of the carotenoid-pigmented plumage patches in 33 *Euplectes* taxa are shown in Figure 1. Objective hue colorimetrics, and discrete color scores derived from these, are listed for *Euplectes* and outgroups in Table 1.

RECONSTRUCTIONS OF COLOR EVOLUTION

Continuous hue

Linear parsimony reconstruction of continuous color hue suggested that extant widowbirds and bishops likely derive from a common ancestor with yellow ($\lambda_{R50} = 522\text{--}536$ nm; Fig. 2) plumage coloration. These results furthermore imply that red coloration (i.e., $\lambda_{R50} > \text{ca. } 555$ nm) has evolved repeatedly and independently in the most recent common ancestor of the “red bishop” clade, and in the ancestors of current *E. axillaris* (fan-tailed widowbird) and *E. progne* (long-tailed widowbird) subspecies. In addition, one or two independent “gains” of red carotenoid coloration, that is, large (>30 nm) increases in hue, has occurred in the *Euplectes* sister group of *Quelea* and *Foudia* (see Appendix S1). Based on midpoints of ranges estimated by linear parsimony, slight decreases in hue (<15 nm) have also occurred in the ancestors of current *E. gierowii* (black bishop), *E. ardens* (red-collared widowbird), and *E. albonotatus* (white-winged widowbird) subspecies.

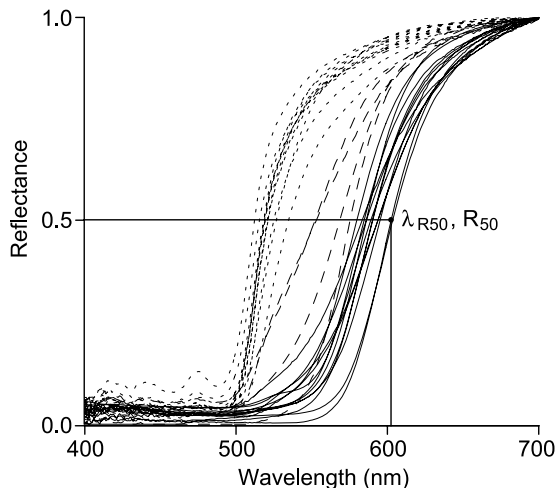


Figure 1. Mean spectral reflectance of the carotenoid-pigmented plumage patches in 33 *Euplectes* taxa. To illustrate differences in spectral position (hue), independent of variation in brightness, reflectance spectra have been normalized to equal maxima. Taxon spectra are sorted into the classes “yellow” (dotted lines), “orange” (dashed) and “red” (solid), based on “k-means” clustering ($k = 3$) of the objective hue colorimetric λ_{R50} (i.e., wavelength of the reflectance midpoint [R_{50}], see Methods, illustrated for the rightmost curve in the graph).

Ancestral reconstruction with squared-change parsimony (Appendix S2) supported a slightly longer wavelength, but still yellow or golden, hue ($\lambda_{R50} = 544$ nm) of the *Euplectes* ancestor. The same three independent origins of red hues were again suggested within the genus, as well as one additional gain in the *Quelea* plus *Foudia* clade. Finally, minor decreases (<15 nm) in color hue were mapped onto branches leading to most yellow groups of the tree, for example, to clades of *Ploceus*, *E. afer* (yellow-crowned bishop), and *E. macrourus* (yellow-mantled widowbird).

Discrete hue

With varying confidence, all analyses of discrete color hue supported the main results derived from continuous reconstructions, that is, a yellow ancestor to extant widowbirds and bishops, and three independent origins of red coloration in the genus (Fig. 2). Inclusion of non-carotenoid pigmented taxa in discrete analyses furthermore showed that complete loss of carotenoid pigmentation has occurred independently in *E. ardens concolor* (the all-black form of red-collared widowbird), the East-African *E. albonotatus eques*, and *E. jacksoni* (Jackson’s widowbird). In addition, likely reversals from red to orange or yellow were suggested to have occurred in the ancestors of *E. gierowii* (black bishop), in *E. ardens ardens* (red-collared widowbird), and, possibly, in *E. albonotatus*.

Reconstructions of three- and four-state hue (Table 1), respectively, were highly concordant, and results are thus only illustrated for the latter, more fine-scaled classification scheme (Fig. 2). The likelihood of yellow ancestors to (1) all *Euplectes*, (2) *Euplectes* minus *E. afer* and *E. aureus*, (3) “true widowbirds” (*E. capensis* to *E. progne*; Fig. 2), (4) and “true widowbirds” minus *E. capensis* and *E. macrourus*, however, received 0.17–0.21 higher likelihood support from reconstruction of three-state hue (see Appendix S2), compared to four-state hue.

Within *Euplectes*, parsimony reconstruction was unambiguous and consistent with maximum likelihood and stochastic character mapping, with two exceptions: First, in the ancestor of the “red bishop” clade (*E. gierowii* to *E. orix*; Fig. 2), the likelihood and posterior probability of red coloration (0.56 and 0.75–0.78, depending on prior settings) was higher than for orange (0.26 and 0.20), which was found to be the most parsimonious state. Second, proportional likelihood support for the most parsimonious state in the common ancestor of *E. axillaris* and *E. albonotatus* (yellow; 0.52) was very similar to that of an alternative state (red: 0.47), and posterior probabilities for these were reversed (0.17–0.19 vs. 0.80–0.82).

Stochastic mappings based on different priors settings for overall transformation rate consistently favored the same ancestral states, although posterior probabilities (integrated over alternative tree topologies) varied somewhat between different prior settings.

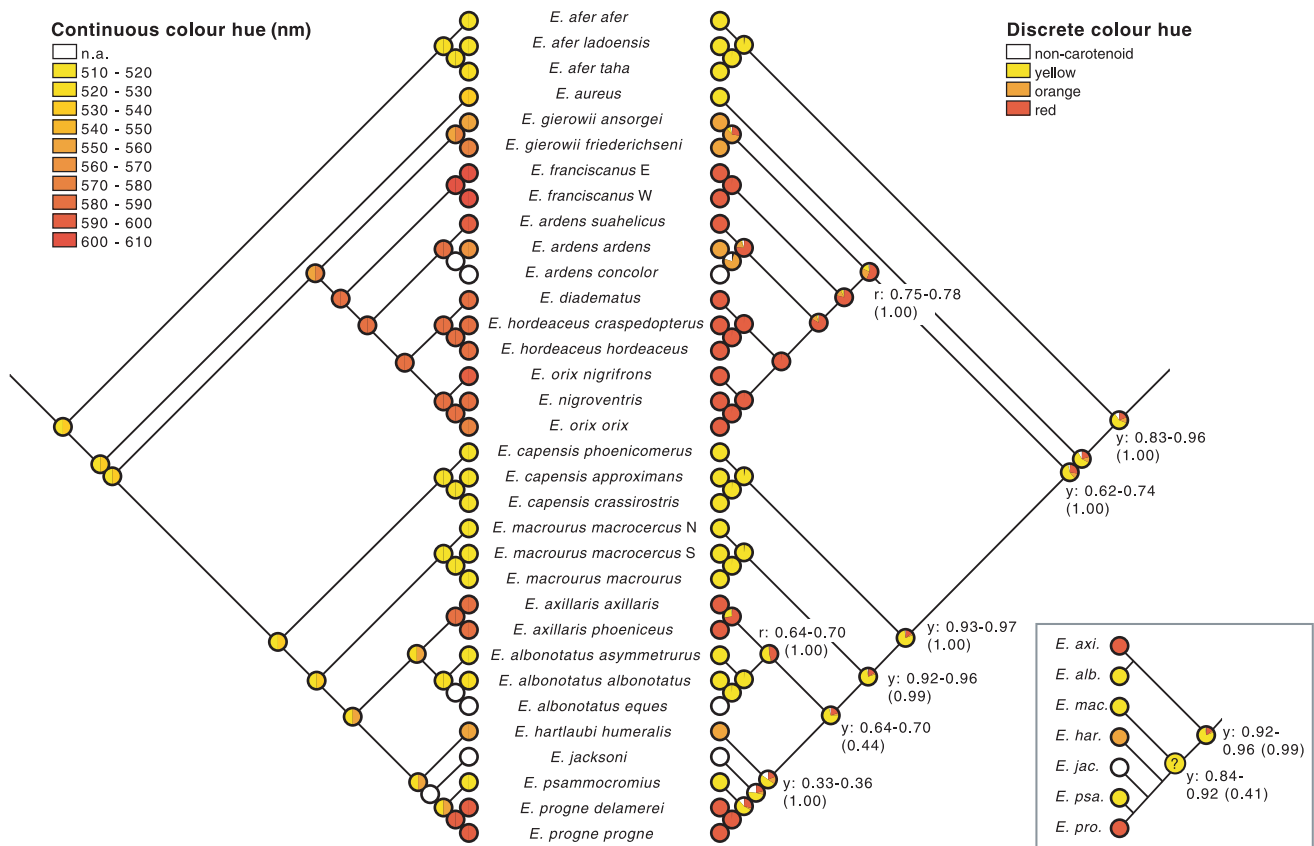


Figure 2. Reconstruction of ancestral carotenoid color hues in *Euplectes*. Circles in the left tree represent linear parsimony estimates of continuous hue states (single, or intervals of equally parsimonious, states). Pie charts in the right tree show proportional likelihoods of alternative discrete hue states. For selected nodes, numbers additionally state the posterior probability of favored states (y = yellow, o = orange, r = red), and of the node itself (in parenthesis; from Prager et al. 2009). The inserted box illustrates an alternative topology for a poorly resolved clade, of importance for conclusions about the homology of red coloration in the group.

Although reconstructions showed considerable uncertainty, the posterior probabilities for yellow ancestors to (1) all *Euplectes* and (2) true widowbirds (see above) were high (0.83–0.96 and 0.93–0.97), suggesting that red coloration has been gained separately in bishops and true widowbirds. Less likely, but implied by lower support for yellow color in the ancestor of *Euplectes* minus *E. afer* and *E. aureus* (0.62–0.74), red color has been lost and subsequently regained in the widowbird clade. Stochastic character mapping on the two alternative topologies within the poorly resolved widowbird clade (Fig. 2 and insert), likewise showed some support (0.64–0.70 or 0.92–0.96) for yellow plumage in the most recent common ancestors of *E. axillaris* and *E. progne*, suggesting independent gains of red coloration in these taxa as well.

Discussion

Ancestral state reconstructions of both discrete and continuous color hue in *Euplectes* suggest a rather directional and convergent evolution of red carotenoid coloration: Extant widowbirds and

bishops derive from a yellow ($\lambda_{R50} = 522\text{--}544$ nm) ancestor, and during the diversification there have been at least two, probably three, convergent gains of red coloration (i.e., $\lambda_{R50} > \text{ca. } 555$ nm), after which there are no strongly supported reversals to yellow and only a single loss of carotenoid coloration (in *E. ardens concolor*). According to the most likely scenario, red hues have evolved independently in the most recent common ancestor of the “red bishop” clade, and in the ancestors of extant *E. axillaris* (fan-tailed widowbird) and *E. progne* (long-tailed widowbird) subspecies.

That the similarity in color hue between “red bishops” (*E. gierowii* to *E. orix*; Fig. 2) and “true widowbirds” (*E. capensis* to *E. progne*) is due to convergent evolution, rather than shared ancestry, is further supported by HPLC analyses of plumage pigments, showing distinct differences in feather pigmentation between the two groups: First, *E. orix* (southern red bishop) and *E. ardens* (red-collared widowbird), both belonging to the “red bishop” clade, achieve orange-red color by adding relatively small amounts of metabolically derived red C4-keto-carotenoids (mainly α -doradexanthin and canthaxanthin) to feathers.

In contrast, the longwave hues of *E. axillaris* (fan-tailed widowbird) derive from deposition of high concentrations of dietary and derived yellow pigments (Andersson et al. 2007; Prager et al. 2009). Importantly, these mechanistic differences support homoplasy in color between these clades, even if their most recent common ancestor (third node from right in right-hand tree, Fig. 2) was not yellow, in which case red coloration has been lost and later regained in the “true widows.” The support for a third independent origin of red plumage color in *E. progne* (long-tailed widowbird) varies considerably with alternative tree topologies (Fig. 2 and insert), but may be resolved by future pigment analyses in this species, and by better phylogenetic resolution in this part of the tree.

RECONSTRUCTION METHODS

Given that evolutionary processes include a stochastic component, ancestral state reconstructions are inevitably uncertain, especially for rapidly evolving traits (Schluter et al. 1997; Losos 1999). Whereas parsimony methods only reflect this uncertainty by displaying equally parsimonious states, maximum likelihood and Bayesian methods provide estimates of accuracy for all reconstructed ancestral states. Following Edwards (1972), Pagel (1999) suggested that the difference in log likelihood between states should be at least two, before one concludes that one is significantly more likely than another. Applying this strict likelihood requirement to color evolution in *Euplectes* would mean that most internal nodes are considered equivocal, at least in terms of four-state hues. However, many of these nodes received close to 0.95, i.e., high, posterior probability in stochastic character mapping. Although the support for reconstructed values varied somewhat between methods, reconstructions of discrete and continuous color gave highly concordant results, regardless of whether parsimony, likelihood, or (Bayesian) stochastic methods were used. This is particularly reassuring because state probabilities in the latter method are integrated over alternative tree topologies, branch lengths, and transformation rates, and because rate priors were intentionally varied within quite wide intervals. Below, we discuss the implications of these results and, for poorly supported nodes, the extent to which alternative reconstructions would lead to different interpretations of the evolutionary history of carotenoid coloration in widowbirds and bishops.

SIGNAL SELECTION AND COSTS

Although multiple gains of a signal selection pressure for red is entirely possible, it is tempting to interpret the convergence of red coloration as a result of a universal and ancestral selection pressure for ever redder and more honest carotenoid displays. As regards the agonistic function, the importance of redness for social status has been established for the phylogenetically quite distant *E. axillaris* and *E. ardens* (Pryke et al. 2001, 2002; Pryke and

Andersson 2003a,b). The generality of red dominance and honesty in other weaverbirds needs further investigation, however, because, for example, red bill color (Shawcross and Slater 1983) but not plumage redness (Dale 2000) was found to be related to individual quality in *Quelea quelea*. Furthermore, color manipulations in a captive *E. orix* population had no consistent effect on social status (Edler and Friedl 2010). Finally, females mated to males with brighter (i.e., likely less carotenoid-pigmented) plumage produced larger clutches in *Foudia madagascariensis* (Estep et al. 2006).

In addition to the convergence of redness, strongly supported by the different underlying mechanisms, ancestral state reconstructions also suggest evolution toward shorter wavelength hues in some branches of the *Euplectes* tree, and even complete loss of carotenoid pigmentation in three taxa: Jackson's widowbird (*E. jacksoni*), the East African subspecies of white-winged widowbird (*E. albonotatus eques*), and the all-black form of the red-collared widowbird (*E. ardens concolor*). These losses seem independent of each other and all are taxa with elongated nuptial tails, exaggerated directionally (Prager and Andersson 2009) in response to female mate choice (Andersson 1982, 1992; Pryke and Andersson 2002, 2005). A trade-off between tail length and carotenoid investment has previously been demonstrated intraspecifically in *E. ardens ardens* (Andersson et al. 2002), but may also contribute to the apparent negative relationship between nuptial tail elongation and coloration in the genus as a whole (S. Andersson et al., unpubl. data).

GENETIC CONSTRAINTS

Given consistent selection for redness and limited variation in ecological constraints (diet, habitat, visual predators), one likely explanation for yellow or no carotenoid coloration in some taxa seems to be genetic or physiological constraints. Indeed, diet manipulation showed that *E. afer* (yellow-crowned bishop) not only lacked red C4-keto-carotenoids (α -doradoxanthin and canthaxanthin) in their feathers, but, unlike *E. orix*, were also unable to manufacture these from yellow dietary precursors (lutein and β -carotene, respectively; Prager et al. 2009). Whether this means that *E. afer*, and presumably other yellow *Euplectes*, lack the actual gene for the hypothesized enzyme, or just its expression (due to, e.g., energetic constraints) remains to be investigated.

IMPLICATIONS FOR SEXUAL SELECTION AND SIGNAL DIVERSIFICATION

As in our reconstruction of male tail elongation (Prager and Andersson 2009), the evolutionary pattern for carotenoid coloration seems to be one of directional and convergent signal exaggeration. There are several reasons why we think this is the expected footprint of sexual selection, despite the many studies indicating lability and divergence (in birds, e.g., Kusmiński et al.

1997; Prum 1997; Kimball et al. 2001; Ödeen and Björklund 2003; Hofmann et al. 2006; Price et al. 2007; Cardoso and Mota 2008):

- We analyze a taxon and signal traits for which sexual selection pressures are uniquely well documented. These signals are furthermore measured in dimensions (i.e., tail length and color hue) that are both targeted by signal receivers, and closely related to honesty-maintaining mechanisms (Andersson 2000; Andersson and Prager 2006). In contrast, when little is known about the details of sexual selection, missing or mismeasuring its targets may invalidate conclusions about its macroevolutionary consequences.
- Younger and less ecologically diverse radiations like this one may present cleaner outcomes of sexually selected signal diversification, not drowning in a multitude of phylogenetically and historically varying constraints. In fact, we think the view of labile and divergent sexual selection partly derives from ascribing reconstructed patterns of signal evolution to only one of several underlying selection processes. Ecological divergence in signal conditions and costs, or conflicts with species recognition, can reverse or redirect signal evolution, but this does not mean that sexual selection per se is an inherently labile and divergent force.
- The distinction between signal selection and constraints highlights the importance of understanding the underlying mechanisms. Without knowing the biochemistry behind red coloration in the two clades (Andersson et al. 2007; Fig. 2), the case for convergence (and perhaps a pre-existing sensory bias) would have relied only on the ancestral state reconstruction and had been much weaker.

CONCLUSION

Conspicuous avian plumage colors have been discussed and intraspecifically tested with respect to social and sexual selection pressures for more than a century. Yet, few investigations have targeted interspecific color diversity by combining data on proximate mechanisms and adaptive functions of coloration in a robust phylogenetic framework. Our reconstruction of the evolution of sexually selected carotenoid coloration in the African widowbirds and bishops (*Euplectes* spp.) provides a valuable perspective on sexually selected signal diversification, and for exploring genetic and physiological constraints on carotenoid coloration in birds. We conclude that extant widowbirds and bishops likely derive from a yellow ancestor, and that redder (longer wavelength) color hues have been repeatedly gained, via different pigment mechanisms. The directional and convergent sexual selection scenario may be more representative than it seems in the literature, but only detected where signal functions and mechanisms are well

known, and social and ecological diversification is limited. These results should guide further experimental research, underlining the important synergy of intra- and interspecific studies for understanding the evolutionary origins of biodiversity.

ACKNOWLEDGMENTS

We are grateful to S. Pryke and J. Örnberg for contributions in the field, to R. Prys-Jones and the Natural History Museum at Tring, UK, for access to specimens, and to J. Pienaar and L. Kvarnemo for comments on the manuscript. This study was supported by grants from the Swedish Research Council (to SA), and (to MP) from Rådman och fru Ernst Collianders Stiftelse, Wilhelm och Martina Lundgrens Vetenskapsfond, and Helge Ax:son Johnsons stiftelse.

LITERATURE CITED

- Akaike, H. 1974. A new look at the statistical model identification. *IEEE Trans. Autom. Control* 19:716–723.
- Andersson, M. 1982. Female choice selects for extreme tail length in a widowbird. *Nature* 299:818–820.
- . 1994. *Sexual selection*. Princeton Univ. Press, Princeton, NJ.
- Andersson, S. 1992. Female preference for long tails in lekking Jackson's widowbirds – experimental evidence. *Anim. Behav.* 43:379–388.
- . 2000. Efficacy and content in avian color signals. Pp. 47–60 in Y. Espmark, T. Amundsen, and G. Rosenqvist, eds. *Animal signals: signalling and signal design in animal communication*. Tapir Academic Press, Trondheim, Norway.
- Andersson, S., and M. Prager. 2006. Quantifying colors. Pp. 41–89 in G. E. Hill and K. J. McGraw, eds. *Bird coloration, vol. 1: mechanisms and measurements*. Harvard Univ. Press, Cambridge, MA.
- Andersson, S., M. Prager, and E. I. A. Johansson. 2007. Carotenoid content and reflectance of yellow and red nuptial plumages in widowbirds (*Euplectes* spp.). *Funct. Ecol.* 21:272–281.
- Andersson, S., S. R. Pryke, J. Örnberg, M. J. Lawes, and M. Andersson. 2002. Multiple receivers, multiple ornaments, and a trade-off between agonistic and epigamic signaling in a widowbird. *Am. Nat.* 160:683–691.
- Badyaev, A. V. 2004. Developmental perspective on the evolution of sexual ornaments. *Evol. Ecol. Res.* 6:975–991.
- Bakker, T. C. M., and P. Sevenster. 1983. Determinants of dominance in male sticklebacks (*Gasterosteus aculeatus* L.). *Behaviour* 86:55–71.
- Blount, J. D., N. B. Metcalfe, T. R. Birkhead, and P. F. Surai. 2003. Carotenoid modulation of immune function and sexual attractiveness in zebra finches. *Science* 300:125–127.
- Bollback, J. P. 2006. SIMMAP: stochastic character mapping of discrete traits on phylogenies. *BMC Bioinform.* 7:88.
- Brockmann, H., and O. Völker. 1934. Der gelbe Federfarbstoff des Kanarienvogels (*Serinus canaria canaria* [L.]) und das Vorkommen von Carotinoiden bei Vögeln. *H.-S. Z. Physiol. Chem.* 224:193–215.
- Brush, A. 1978. Avian pigmentation. Pp. 141–164 in M. Florin, T. S. Bradley, and A. Brush, eds. *Chemical zoology*. Academic Press, New York.
- Cardoso, G. C., and P. G. Mota. 2008. Speciation evolution of coloration in the genus *Carduelis*. *Evolution* 62, 753–762.
- Crook, J. H. 1964. The evolution of social organisation and visual communication in the weaverbirds (Ploceinae). *Behav. Suppl.* 10:1–178.
- Dale, J. 2000. Ornamental plumage does not signal male quality in red-billed queleas. *Proc. R. Soc. B* 267:2143–2149.
- Edler, A. U., and T. W. P. Friedl. 2010. Plumage colouration, age, testosterone and dominance in male red bishops (*Euplectes orix*): A laboratory experiment. *Ethology* 116:1–15.

- Edwards, A. W. F. 1972. Likelihood. Cambridge Univ. Press, Cambridge, U.K.
- Endler, J. A. 1980. Natural selection on color patterns in *Poecilia reticulata*. *Evolution* 34:76–91.
- Endler, J., and A. Basolo. 1998. Sensory ecology, receiver biases and sexual selection. *Trends Ecol. Evol.* 13:415–420.
- Estep, L. K., M. D. Shawkey, and G. E. Hill. 2006. Carotenoid-based breast plumage color, body condition and clutch size in red fodies (*Foudia madagascariensis*). *Ostrich* 77:164–169.
- Evans, M. R., and B. J. Hatchwell. 1992. An experimental study of male adornment in the scarlet-tufted malachite sunbird: I. The role of pectoral tufts in territorial defence. *Behav. Ecol. Sociobiol.* 29:413–419.
- Evans, M. R., and K. Norris. 1996. The importance of carotenoids in signaling during aggressive interactions between male firemouth cichlids (*Cichlasoma meeki*). *Behav. Ecol.* 7:1–6.
- Farris, J. S. 1970. Methods of computing Wagner trees. *Syst. Zool.* 19:83–92.
- Fox, H. M., and G. Vevers. 1960. The nature of animal colors. Macmillan Co., New York.
- Goodwin, T. W. 1984. The biochemistry of carotenoids, vol. II: animals. Chapman & Hall, New York.
- Hamilton, W. D., and M. Zuk. 1982. Heritable true fitness and bright birds: a role for parasites? *Science* 218:384–387.
- Hansen, A. J., and S. Rohwer. 1986. Coverable badges and resource defence in birds. *Anim. Behav.* 34:69–76.
- Hartigan, J. A., and M. A. Wong. 1979. Algorithm AS 136: a k-means clustering algorithm. *J. R. Stat. Soc. Ser. C.* 28:100–108.
- Hartley, R. C., and M. W. Kennedy. 2004. Are carotenoids a red herring in sexual display? *Trends Ecol. Evol.* 19:353–354.
- Harvey, P. H., and M. D. Pagel. 1991. The comparative method in evolutionary biology. Oxford Univ. Press, Oxford, U.K.
- Hill, G. E. 1990. Female house finches prefer colorful males: sexual selection for a condition-dependent trait. *Anim. Behav.* 40:563–572.
- . 1994. Trait elaboration via adaptive mate choice: sexual conflict in the evolution of signals of male quality. *Ethol. Ecol. Evol.* 6:351–370.
- . 1996. Redness as a measure of the production cost of ornamental coloration. *Ethol. Ecol. Evol.* 8:157–175.
- Hill, G. E., and K. J. McGraw, eds. 2006a. Bird coloration, vol 2. Function and evolution. Harvard Univ. Press, Cambridge, MA.
- . 2006b. Bird coloration, vol 1. Mechanisms and measurements. Harvard Univ. Press, Cambridge, MA.
- Hofmann, C. M., T. W. Cronin, and K. E. Omland. 2006. Using spectral data to reconstruct evolutionary changes in coloration: carotenoid color evolution in new world orioles. *Evolution* 60:1680–1691.
- Hudon, J. 1991. Unusual carotenoid use by the Western Tanager (*Piranga ludoviciana*) and its evolutionary implications. *Can. J. Zool.-Rev. Can. Zool.* 69:2311–2320.
- Huelsenbeck, J. P., R. Nielsen, and J. P. Bollback. 2003. Stochastic mapping of morphological characters. *Syst. Biol.* 52:131–158.
- Huey, R. B., and A. F. Bennett. 1987. Phylogenetic studies of coadaptation: preferred temperatures versus optimal performance temperatures of lizards. *Evolution* 41:1098–1115.
- Kiere, L. M., C. M. Hofmann, J. J. Price, T. W. Cronin, and K. E. Omland. 2009. Discrete evolutionary color changes in caciques suggest different modes of carotenoid evolution between closely related taxa. *J. Avian Biol.* 40:605–613.
- Kimball, R. T., E. L. Braun, J. D. Ligon, V. Lucchini, and E. Randi. 2001. A molecular phylogeny of the peacock-pheasants (Galliformes: *Polyplectron* spp.) indicates loss and reduction of ornamental traits and display behaviours. *Biol. J. Linn. Soc.* 73:187–198.
- Kritzler, H. 1943. Carotenoids in the display and eclipse plumages of bishop birds. *Physiol. Zool.* 15:241–255.
- Kusmiński, R., G. Borgia, A. Uy, and R. H. Crozier. 1997. Labile evolution of display traits in bowerbirds indicates reduced effects of phylogenetic constraint. *Proc. R. Soc. B* 264:307–313.
- Lee, C., S. Blay, A. Ø. Mooers, A. Singh, and T. H. Oakley. 2006. CoMET: a Mesquite package for comparing models of continuous character evolution on phylogenies. *Evol. Bioinform. Online* 2:193–196.
- Lewis, P. O. 2001. A likelihood approach to estimating phylogeny from discrete morphological character data. *Syst. Biol.* 50:913–925.
- Losos, J. B. 1999. Uncertainty in the reconstruction of ancestral character states and limitations on the use of phylogenetic comparative methods. *Anim. Behav.* 58:1319–1324.
- Lozano, G. A. 1994. Carotenoids, parasites, and sexual selection. *Oikos* 70:309–311.
- Maddison, W. P., and D. R. Maddison. 2004. Mesquite: a modular system for evolutionary analysis. Version 2.71 Available at: <http://mesquiteproject.org>.
- McGraw, K. J. 2006. The mechanics of carotenoid-based coloration in birds. Pp. 177–242 in G. E. Hill and K. J. McGraw, eds. Bird coloration. Harvard Univ. Press, Cambridge, MA.
- Milinski, M., and C. M. Bakker. 1990. Female sticklebacks use male coloration in mate choice and hence avoid parasitized males. *Nature* 344:330–333.
- Møller, A. P., C. Biard, J. D. Blount, D. C. Houston, P. Ninni, N. Saino, and P. F. Surai. 2000. Carotenoid-dependent signals: indicators of foraging efficiency, immunocompetence or detoxification ability? *Avian Poult. Biol. Rev.* 11:137–159.
- Nielsen, R. 2002. Mapping mutations on phylogenies. *Syst. Biol.* 51:729–739.
- Oakley, T. H., Z. L. Gu, E. Abouheif, N. H. Patel, and W. H. Li. 2005. Comparative methods for the analysis of gene-expression evolution: an example using yeast functional genomic data. *Mol. Biol. Evol.* 22:40–50.
- Ödeen, A., and M. Björklund. 2003. Dynamics in the evolution of sexual traits: losses and gains, radiation and convergence in yellow wagtails (*Motacilla flava*). *Mol. Ecol.* 12:2113–2130.
- Olson, V. A., and I. P. F. Owens. 1998. Costly sexual signals: are carotenoids rare, risky or required? *Trends Ecol. Evol.* 13:510–514.
- Omland, K. E., and C. M. Hofmann. 2006. Adding color to the past: ancestral-state reconstruction of coloration. Pp. 417–454 in G. E. Hill and K. J. McGraw, eds. Bird coloration, vol. 2: function and evolution. Harvard Univ. Press, Cambridge, MA.
- Omland, K. E., and S. M. Lanyon. 2000. Reconstructing plumage evolution in orioles (*Icterus*): repeated convergence and reversal in patterns. *Evolution* 54:2119–2133.
- Pagel, M. 1999. The maximum likelihood approach to reconstructing ancestral character states of discrete characters on phylogenies. *Syst. Biol.* 48:612–622.
- Prager, M., and S. Andersson. 2009. Phylogeny and evolution of sexually selected tail ornamentation in widowbirds and bishops (*Euplectes* spp.). *J. Evol. Biol.* 22:2068–2076.
- Prager, M., E. I. A. Johansson, and S. Andersson. 2008. A molecular phylogeny of the African widowbirds and bishops, *Euplectes* spp. (Aves: Passeridae: Ploceinae). *Mol. Phylogenet. Evol.* 46:290–302.
- . 2009. Differential ability of carotenoid C4-oxygenation in yellow and red bishop species (*Euplectes* spp.) *Comp. Biochem. Physiol. Part B Biochem. Mol. Biol.* 154:373–380.
- Price, J. J., N. R. Friedman, and K. E. Omland. 2007. Song and plumage evolution in the new world orioles (*Icterus*) show similar lability and convergence in patterns. *Evolution* 61:850–863.
- Prum, R. O. 1997. Phylogenetic tests of alternative intersexual selection mechanisms: trait macroevolution in a polygynous clade (Aves: Pipridae). *Am. Nat.* 149:668–692.
- Pryke, S. R. 2007. Fiery red heads: female dominance among head color morphs in the Gouldian finch. *Behav. Ecol.* 18:621–627.

- Pryke, S. R., and S. Andersson. 2002. A generalized female bias for long tails in a short-tailed widowbird. *Proc. R. Soc. Biol. Sci. Ser. B* 269:2141–2146.
- . 2003a. Carotenoid-based epaulettes reveal male competitive ability: experiments with resident and floater red-shouldered widowbirds. *Anim. Behav.* 66:217–224.
- . 2003b. Carotenoid-based status signalling in red-shouldered widowbirds (*Euplectes axillaris*): epaulet size and redness affect captive and territorial competition. *Behav. Ecol. Sociobiol.* 53:393–401.
- . 2005. Experimental evidence for female choice and energetic costs of male tail elongation in red-collared widowbirds. *Biol. J. Linn. Soc.* 86:35–43.
- Pryke, S. R., S. Andersson, M. J. Lawes, and S. E. Piper. 2002. Carotenoid status signaling in captive and wild red-collared widowbirds: independent effects of badge size and color. *Behav. Ecol.* 13:622–631.
- Pryke, S. R., and S. C. Griffith. 2006. Red dominates black: agonistic signalling among head morphs in the color polymorphic Gouldian finch. *Proc. R. Soc. B* 273:949–957.
- Pryke, S. R., M. J. Lawes, and S. Andersson. 2001. Agonistic carotenoid signalling in male red-collared widowbirds: aggression related to the color signal of both the territory owner and model intruder. *Anim. Behav.* 62:695–704.
- R Development Core Team. 2009. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.
- Rogers, J. S. 1984. Deriving phylogenetic trees from allele frequencies. *Syst. Zool.* 33:52–63.
- Schluter D., and T. Price. 1993. Honesty, perception and population divergence in sexually selected traits. *Proc. R. Soc. B* 253:117–122.
- Schluter, D., T. Price, A. O. Mooers, and D. Ludwig. 1997. Likelihood of ancestor states in adaptive radiation. *Evolution* 51:1699–1711.
- Shawcross, J. E., and P. J. B. Slater. 1983. Agonistic experience and individual recognition in male *Quelea quelea*. *Behav. Process.* 9:49–60.
- Stradi, R. 1998. The color of flight. Solei Gruppo Editoriale Informatico, Milan, Italy.
- Swofford, D. L., and W. P. Maddison. 1987. Reconstructing ancestral character states under Wagner parsimony. *Math. Biosci.* 87:199–229.
- Thompson, C. W., N. Hillgarth, M. Leu, and H. E. McClure. 1997. High parasite load in house finches (*Carpodacus mexicanus*) is correlated with reduced expression of a sexually selected trait. *Am. Nat.* 149:270–294.
- von Schantz, T., S. Bensch, M. Grahn, D. Hasselquist, and H. Wittzell. 1999. Good genes, oxidative stress and condition-dependent sexual signals. *Proc. R. Soc. B* 266:1–12.
- Wang, I. J., and H. B. Shaffer. 2008. Rapid color evolution in an aposematic species: a phylogenetic analysis of color variation in the strikingly polymorphic strawberry poison-dart frog. *Evolution* 62:2742–2759.
- Wiens, J. J. 2001. Widespread loss of sexually selected traits: how the peacock lost its spots. *Trends Ecol. Evol.* 16:517–523.
- Wolfenbarger, L. L. 1999. Red coloration of male northern cardinals correlates with mate quality and territory quality. *Behav. Ecol.* 10:80–90.
- Zahn, S. N., and S. I. Rothstein. 1999. Recent increase in male house pinch plumage variation and its possible relationship to avian pox disease. *Auk* 116:35–44.

Associate Editor: M. Webster

Supporting Information

The following supporting information is available for this article:

Appendix S1. Reconstruction of ancestral carotenoid color hues in the *Euplectes* and outgroups (*Amblyospiza*, *Ploceus*, *Foudia*, and *Quelea*). Circles in the left tree represent linear parsimony estimates of continuous hue states (single, or intervals of equally parsimonious, states). Pie charts in the right tree show proportional likelihoods of alternative discrete (four-state) hue.

Appendix S2. Reconstruction of ancestral carotenoid color hues in the *Euplectes* and outgroups (*Amblyospiza*, *Ploceus*, *Foudia*, and *Quelea*). Circles in the left tree represent squared-change parsimony estimates of continuous hue states. Pie charts in the right tree show proportional likelihoods of alternative discrete (three-state) hue.

Supporting Information may be found in the online version of this article.

Please note: Wiley-Blackwell is not responsible for the content or functionality of any supporting information supplied by the authors. Any queries (other than missing material) should be directed to the corresponding author for the article.